Received: 2012.12.18 Accepted: 2013.04.30 Published: 2013.05.24	Epstein-Barr virus-associated lymphoproliferative disorders* Choroby limfoproliferacyjne związane z zakażeniem wirusem Epsteina-Barr		
	Ewelina Grywalska¹, Justyna Markowicz¹, Piotr Grabarczyk², Marcin Pasiarski³ Jacek Roliński¹		
	¹ Department of Clinical Immunology and Immunotherapy, Medical University of Lublin, Lublin, Poland ² Department of Immunohaematology, Institute of Haematology and Transfusion Medicine, Warsaw, Poland ³ Department of Hematology, Hollycross Cancer Center, Kielce, Poland		
	Summary		
	The Epstein–Barr virus (EBV) is one of the most common human viruses, infecting more than 90% of the world's adult population. In some individuals the interplay between EBV replication, latency and immune control can be disrupted and evokes prolonged proliferation of EBV-infected lymphocytes and their malignant transformation. Since its discovery as the first human tumor virus, EBV has been implicated in the development of a wide range of human cancers. The evidence for an association with EBV is the strongest for Burkitt's lymphoma, NK/T cell lymphoma, nasopharyngeal carcinoma, Hodgkin's lymphoma and for malignant lymphomas in immune incompetent patients. Additionally, certain epithelial cell tumors, such as gastric carcinoma and breast carcinoma, have been defined as EBV related. However, the virus may be encountered in other types of malignancies. The oncogenic potential of EBV is related to its ability to infect and transform B lymphocytes into continuously growing lymphoblastoid cell lines. EBV encodes a series of products mimicking several growth, transcription and anti-apoptotic factors, to usurp control of the pathways that regulate diverse homeostatic cellular functions. However, the exact mechanism by which EBV promotes oncogenesis remains unclear. The focus of this review is to summarize the current knowledge of oncogenic potential of the Epstein–Barr virus and its role in the pathogenesis of EBV-associated lymphoproliferative disorders.		
Keywords:	EBV • lymphoma • lymphoproliferative disorder • oncogenesis • tumor		
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Author's address:	Ewelina Grywalska, M.D., Ph. D. Department of Clinical Immunology and Immunotherapy, Medical Uni-		

Author's address: Ewelina Grywalska, M.D., Ph. D. Department of Clinical Immunology and Immunotherapy, Medical University of Lublin, 4a Chodźki Street, 20-093 Lublin, Poland; e-mail: ewelina.grywalska@gmail.com

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List of abbreviations:

Ag – antigen; AID – activation-induced cytidine deaminase; AIDS – acquired immune deficiency syndrome; AITL – angioimmunoblastic T-cell lymphoma; APC – antigen-presenting cell; BCR - B-cell receptor; BL - Burkitt's lymphoma; CAEBV - chronic active Epstein-Barr virus disease; CD – cluster of differentiation; CD40L – CD40 ligand; CMV – cytomegalovirus; CTL – cytotoxic T lymphocyte; DC – dendritic cell; DLBCL – diffuse large B-cell lymphoma; EA – early antigen; EBERs - EBV-encoded small RNAs; EBNA - Epstein-Barr nuclear antigen; EBV - Epstein-Barr virus; GC – germinal center; HHV – human herpes virus; HIV – human immunodeficiency virus; HL – Hodgkin's lymphoma; HLA – human leukocyte antigens; HRS – Hodgkin and Reed-Sternberg cells; HSV – herpes simplex virus; HSCT – hematopoietic stem cell transplantation; HTLV-1 – human T-cell leukemia virus-1; IFN – interferon; Ig – immunoglobulin; IL – interleukin; KSHV – Kaposi's sarcoma herpes virus; LCL - lymphoblastoid cell line; LD - lymphocyte depleted; LMP - latent membrane protein; LP – lymphocyte predominant; LPDs – lymphoproliferative disorders; MALT - mucosa-associated lymphoid tissue; MC - mixed cellularity; MHC - major histocompatibility complex; NHL - non-Hodgkin's lymphoma; NK - natural killer; NPC - nasopharyngeal carcinoma; NS - nodular sclerosing; PBMC - peripheral blood mononuclear cells; PCNSL - primary central nervous system lymphoma; PEL – primary effusion lymphoma; PLT – platelets; PTLD – post-transplant lymphoproliferative disease; RBC - red blood cells; RNA - ribonucleic acid; SCID - severe combined immunodeficiency; TAA - tumor-associated antigens; TCR - T-cell receptor; TGF transforming growth factor; Th – T helper; TLR – Toll-like receptor; TNF – tumor necrosis factor; VCA – viral capsid antigen

INTRODUCTION

Epstein-Barr virus (EBV), also called human herpes virus 4 (HHV-4), was the first human virus to be directly implicated in carcinogenesis. EBV is a part of the Herpesviridae family [99,117]. It was discovered by Michael Epstein and Yvonne Barr in 1964 during their research on Burkitt's lymphoma (BL) [32]. Since its discovery, EBV has been found in a variety of other tumor types. The evidence for an association with EBV is the strongest for BL, NK/T-cell lymphoma, nasopharyngeal carcinoma (NPC), Hodgkin's lymphoma (HL) and for malignant lymphomas in immune incompetent patients [47]. Additionally, certain epithelial cell tumors, such as gastric carcinoma [107] and breast carcinoma [5], have been found to be EBV related [78]. However, the virus may be encountered in other types of malignancies.

Like other members of the Herpesviridae, EBV virions have a double-stranded, linear DNA genome encoding approximately 100 genes surrounded by a protein capsid. A protein tegument lies between the capsid and the envelope. Mature virions are 120-180 nm in diameter. EBV infects more than 90% of the world's adult population. Infection usually occurs early in childhood. EBV infections are most prevalent in developing countries, in populations of low socioeconomic status [42, 116]. In countries with stringent hygiene practices, EBV seroprevalence tends to increase gradually with age, showing two seroconversion peaks: at 2 to 4 years and at 14 to 18 years [41]. The mean seroprevalence in children is approximately 50% and increases steadily to a value of 90% to 99% in adults [4].

There are two major strains of EBV (type 1 and type 2), differing in organization of the genes that encode the EBV nuclear antigen (EBNA) [90]. Both types are detected all over the world, with type 1 being the most frequent,

although in some regions (e.g. central Africa, Papua New Guinea and Alaska) type 2 definitely predominates [37, 94]. The virus is passed to uninfected individuals with saliva or spread through close oral contact, but transmission by transfusion has also been documented [7].

In spite of frequent virus detection in latently infected blood donors, transmission of EBV infection by transfusion is thought to be relatively infrequent. This phenomenon could be explained by the fact that most adult recipients of blood and blood products are already immune to EBV. Moreover, blood from seropositive donors contains EBV-neutralizing antibodies and specific memory cells, which may protect the recipient from infection. Although we do not know exact EBV infectivity by blood and its components, it seems that the viral load in blood from healthy seropositive donors, which is normally low (5/106-1/107 peripheral blood mononuclear cells), is rather below an infection dose. In addition, it was established that the viability of B lymphocytes carrying the EBV genome may decline during blood storage. The risk of EBV transmission from red blood cell (RBC) and/or platelet (PLT) transfusions is also significantly reduced by leukoreduction. Thus, in most instances, EBV genomes contained in blood products should not cause severe disease when the transfused recipient is immune competent. However, whole blood and blood components as a potential source of infection should be kept in mind as an association between transfusion and EBV infection, especially in immunosuppressed young patients, is still being discussed [104].

Most primary EBV infections in normal individuals are unapparent, but occasionally EBV can cause acute infectious mononucleosis, which is a self-limited disease. Following primary EBV infection, individuals remain lifelong carriers of the virus. EBV then persists latently in the host within long-life memory B cells [73]. The growth of B cells latently infected with EBV is normally controlled by the host immune response, particularly by virus-specific T cells [41]. However, in some individuals, the virus is implicated in the development of malignancy [25]. It is well documented that in the immunocompromised hosts, the interplay between EBV replication, latency and immune control can be disrupted and evokes prolonged proliferation of EBV-infected lymphocytes and their malignant transformation [17, 34]. There are hypotheses that there is an association between EBV infection and autoimmune [31, 111] and allergic diseases [61,97].

EBV can infect both B cells [9] and epithelial cells [100]. The role of epithelial cells in the life cycle of EBV is still incompletely defined. It is generally believed that epithelial cells in the oropharynx represent the site of primary EBV infection and replication [10]. This is followed by a latent infection of B lymphocytes. The B-lymphotropic nature of EBV is evidenced by the ability to immortalize normal resting B lymphocytes in vitro, converting them into permanently growing lymphoblastoid cell lines (LCL) [93]. EBV-infected B lymphocytes carry the viral genome in a latent form. In some circumstances, EBV latency may reactivate, resulting in the expression of viral genes encoding series of products stimulating anti-apoptotic molecules, cytokines, and signal transducers [77]. Disrupted control of the cellular pathways regulating a wide variety of homeostatic cellular functions leads to neoplastic transformation.

In healthy individuals, EBV is latently maintained in memory B cells, which express only the transcripts for EBV small RNAs (EBERs). This state is termed latency 0 and allows for persistence of the virus in a way that is nonpathogenic and not detectable by the immune system [103].

Three types of latent genes have been described (Figure 1). Type I latency, characterized by the expression of Epstein–Barr nuclear antigen 1 (EBNA-1) and two small noncoding Epstein–Barr RNAs (EBERs), was found in BL [19]. EBV gene expression in latency II, associated with classic HL and T-cell non-Hodgkin's lymphoma [91], is limited to EBNA-1, EBERs, latent membrane protein (LMP)-1, LMP-2A and LMP-2B. Latency III usually involves the unrestricted expression of all EBNAs, EBERs, and LMPs [22] and occurs mainly in immunocompromised individuals suffering from post-transplant lymphoproliferative disorders (PTLDs), HIV-associated lymphoproliferative disorders and in lymphoblastoid cell lines [117].

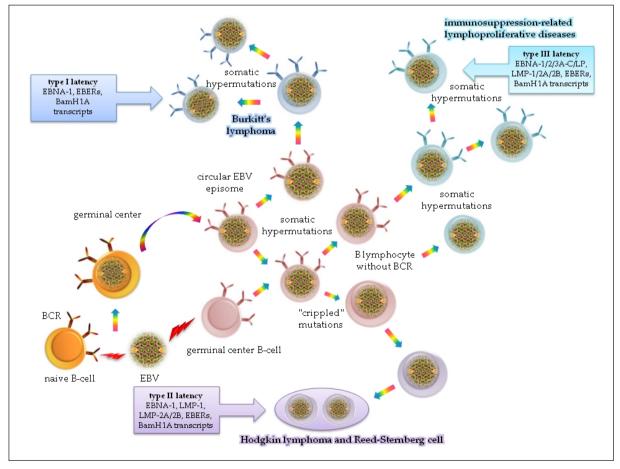


Fig. 1. Three forms of Epstein-Barr virus latency

The ability of EBV to efficiently transform B cells in culture to immortalized, transformed cells undoubtedly accentuates its connection to human cancers [83]. Although EBV may be essential for tumorigenesis, it is not generally sufficient on its own. Other factors such as specific failure of immune recognition, stimulation of B-cell proliferation by other infections, appearance of secondary genetic aberrations or mutations may also be important [99].

EBV usually infects B cells via the B-cell specific CD21 antigen [35] and EBV-associated hematologic malignancies are predominantly of B cell type. In addition, EBV is associated with rare T- and NK-cell malignancies [53,59]. EBV-positive lymphomas can be divided into those occurring in immunodeficient individuals, which are true virally driven lymphomas, such as PTLD and HIV-associated immunoblastic lymphoma, and those occurring in immunocompetent individuals.

EBV-ASSOCIATED LYMPHOMAS IN IMMUNOCOMPETENT INDIVIDUALS

Burkitt's lymphoma

BL is an aggressive lymphoma associated with EBV infection. It is classified as a non-Hodgkin's lymphoma (NHL) and has the fastest doubling time among human tumors [27]. Based on clinical and epidemiological characteristics BL is subdivided into three categories: endemic BL (eBL), sporadic BL (sBL), and HIV-associated BL. Studies suggest that variations differ in geographical distribution and degree of association with EBV. About 95% of eBL cases are associated with EBV and are commonly found in equatorial Africa and Papua New Guinea [105, 118]. In contrast, only 5-15% of sBL, affecting children and young adults, throughout the world and 40% of HIV-associated BL are EBV positive [115]. Subtypes of BL also differ in clinical manifestation. Typically, eBL presents as tumors affecting the jaw and facial bones, while sBL more commonly arises in the gut and upper respiratory tract, forming tumors in the Waldeyer ring [13, 119]. HIV-associated BL characteristically involves the lymph nodes and bone marrow [12]. For all three types of BL, males are more commonly affected than females [101].

The mechanism by which EBV contributes to the development of Burkitt's lymphoma is not entirely understood. The EBV associated BL demonstrates that the complex interaction of EBV with B cells predisposes to BL development [74, 101]. Malaria infection in endemic regions is considered as another co-factor for the development of BL [16]. It is assumed that hyperstimulation of B cells and suppression of T-cell activity by malaria allow for reactivation of EBV in infected B cells, which consequently increase in numbers. The alternative explanation is that malaria infection compromises EBV-specific immune control, leading to immune escape of an EBV-infected B cell including those in which a c-myc translocation has occurred [11]. In all variations of BL, constitutive activation of the c-myc oncogene through its translocation into one of the immunoglobulin loci is clearly the key factor of the oncogenesis [118]. 80% of all BL shows a t(8:14) translocation; the other observed translocations are t(8:22) and t(8:2) [3, 30]. The translocation is dependent on the enzyme activity of AID (activation-induced cytidine deaminase) [44, 80]. AID is highly expressed in the germinal center (GC) to provide the class switch and hypermutation of the Ig variable region. After translocation, the activated myc can lead to cell growth and proliferation; however, it also leads to apoptosis in the absence of apoptosis-inhibiting signals. These signals may be provided by EBV proteins (EBNA-1) [8, 76]. Some studies have shown BL cases that do not show any MYC translocation, yet still overexpress c-myc. This has been demonstrated for <10% of sBL and linked to miRNA deregulation [1].

Most EBV-positive cases exhibit a restrictive pattern of expression of latent encoded proteins, only expressing EBNA-1 and the EBERs (latency I) [118]. Because EBNA-1 protein is poorly antigenic and has little to no HLA class I response, the CD8+ T-cell response to BL is largely diminished. Various pathways by which BL escapes immune detection by inhibiting both HLA class I- and II-mediated Ag presentation to T cells are essential to the disease pathogenesis [36,72].

It was recently reported that some cases, in addition to EBNA-1 and the EBERs, express EBNA-3A, EBNA-3B, EBNA-3C, and EBNA leader protein but still lack EBNA-2 and the latent membrane proteins [60]. EBNA-1 plays a crucial role in the maintenance and replication of the viral genome, but its oncogenic potential is highly controversial [114,118]. Conversely, as the EBERs are believed to possess anti-apoptotic activity, it has been postulated that they may play an essential role in the oncogenesis of BL [62].

Hodgkin's lymphoma

Hodgkin's lymphoma accounts for about 1% of all cancers and 30% of lymphoid malignancies worldwide [43]. Epidemiologic studies of HL demonstrate a remarkable diversity of the incidence according to age, sex, ethnicity, geographic location and socioeconomic status [21,51]. According to the WHO classification, HL can be histologically divided into four subtypes: lymphocyte predominant (LP), nodular sclerosing (NS), mixed cellularity (MC) and lymphocyte depleted (LD) [38]. Not all subtypes harbor EBV to the same degree. EBV positivity in lymphoma tissue is discerned in 70% of MC Hodgkin's disease, 95% of LD Hodgkin's disease, and 10–40% of NS. LP Hodgkin's disease subtype is almost always EBV negative [24].

The role that EBV plays in HL is still not fully understood. Several lines of evidence including increased risk in individuals with a past history of infectious

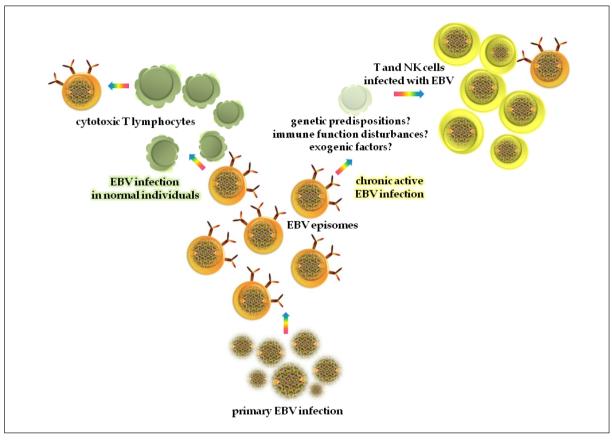


Fig. 2. Pathogenesis of CAEBV

mononucleosis [2], elevated antibody titers to EBV viral capsid antigen [71] and demonstration of the virus in the malignant cells [110] link EBV to HL. HL is characterized by an expansion of Hodgkin and Reed-Sternberg (HRS) cells representing transformed B cells and constituting a minority of the tumor mass [67]. The feature of HRS cells is constant activation of antiapoptotic transcription factor NF- κ B, essential for HRS cell survival [85].

EBV-positive HL has expression of EBNA-1, LMP-1, LMP-2 and the EBERs, indicative of type 2 latency [28]. HRS cells often have destructive immunoglobulin variable (V) gene mutation and therefore lose expression of the B-cell receptor (BCR). Germinal-centre B cells that acquire such mutations are normally eliminated by apoptosis in the germinal centre. The current scenario for HL is that the expression of latent membrane protein 1 (LMP-1) and LMP-2A may prevent apoptosis by mimicking CD40 and BCR signaling, respectively. [57, 65]. Furthermore, to evoke its own survival, HRS cells employ several mechanisms, which are able to suppress a supportive microenvironment of immune and stromal cells and local immune responsiveness [33]. HRS cells have been shown to produce immunosuppressive cytokines such as IL-10, IL-13 and TGF- β [39,45,58,72] as one of the elements of immune escape in Hodgkin's lymphoma.

T-cell lymphomas

In contrast to its role in B-lymphomagenesis, EBV has only incidentally been associated with T-cell lymphomas. Several types of non-B-cell non-Hodgkin's lymphoma are associated with EBV [48, 52]. We will focus on angioimmunoblastic and nasal T/NK cell lymphoma.

Angioimmunoblastic T-cell lymphoma (AITL) is a rare neoplasm but represents the most common subtype of peripheral T-cell lymphomas [50]. Viral genome is detected in up to 100% of AITL lymph nodes [109]. However, EBV can be detected only in B cells [14]. The presence of EBV in only a subpopulation of cells suggests that EBV infection is secondary to malignancy or that the viral genome has been lost from the malignant cell [99]. AITL is characterized by systemic disease, a polymorphous infiltrate primarily involving lymph nodes, proliferation of high endothelial venules and follicular dendritic cells [113]. Clinically it occurs as a widespread lymphadenopathy, extranodal disease, immune-mediated hemolysis, and polyclonal hypergammaglobulinemia, and presents poor prognosis [46].

Nasal-type T/NK-cell lymphoma is a rare tumor, almost always associated with EBV, prevalent in Asia and especially in China [49]. Nasal T/NK lymphoma cells exhibit several unique genotypic and phenotypic features. These features include an absence of T-cell antigens, the expression of the NK cell marker CD56, and the absence of T-cell receptor gene rearrangement [112]. The nasal region is the most frequent site of involvement but the tumor may also appear at other extranodal sites such as skin, testis, kidney, upper gastrointestinal tract, and the orbit [23, 88].

Some nasal NK/T-cell lymphomas develop from longlasting EBV infection termed chronic active EBV infection (CAEBV). CAEBV is an EBV-associated syndrome characterized by a variety of serious hematological disorders, including malignant lymphoma. EBV was found to infect circulating T and/or NK cells in patients with CAEBV. These EBV-infected cells express EBNA-1, LMP-1, and LMP-2A, a type II form of EBV latency, which is also observed in nasopharyngeal carcinoma (NPC), HL, and peripheral T-cell lymphoma. CAEBV may thus represent a subset of EBV-associated T- and/or NK-cell lymphoproliferative disorders [56]. Most lymphomas associated with CAEBV have been reported to be nodal T-cell [55, 82] but in some cases extranodal T-cell lymphoma was diagnosed in the skin, kidneys, spleen, pancreas, and meninges [6]. Figure 2 presents the pathogenesis of CAEBV.

EBV-ASSOCIATED LYMPHOMAS IN IMMUNOCOMPROMISED INDIVIDUALS

Patients with primary immunodeficiencies such as Wiskott-Aldrich syndrome, severe combined immunodeficiency (SCID) or X-linked lymphoproliferative disease are prone to develop EBV-related lymphoproliferative disorders (LPDs). However, EBV-related LPDs are more frequent in patients with secondary immunodeficiencies with AIDS or in organ transplant recipients who receive immunosuppressive therapy [26].

HIV-associated lymphoproliferative disorders

HIV-associated lymphoproliferative disorders represent a heterogeneous group of diseases, arising in the presence of HIV-associated immunodeficiency. According to the WHO classification, HIV-associated lymphomas are categorized into: (a) lymphomas also occurring in immunocompetent patients, including BL, diffuse large B-cell lymphoma (DLBCL) with centroblastic/immunoblastic features, extranodal marginal zone lymphoma of MALT type, peripheral T-cell lymphoma and HL; (b) lymphomas occurring more specifically in HIV-positive patients, including primary effusion lymphoma and plasmablastic lymphoma of the oral cavity; (c) lymphomas also occurring in other immunodeficiency states, including polymorphic B-cell lymphoma (PTLD-like) [84]. The increased risk for lymphoma among HIV-infected individuals appears to be related to multiple factors, including duration and degree of immunosuppression, induction of cytokines leading to B-cell proliferation, and opportunistic infections with oncogenic herpesviruses such as EBV and HHV-8 [64]. The evidence of EBV involvement in development of HIV-related lymphomas is the demonstration of monoclonal virus in the tumors. EBV is present in approximately 60% of HIV-related lymphomas, but the proportion of virus-positive tumors varies depending on histological subtype [26].

BLs make up the largest group of HIV-associated non-Hodgkin lymphomas, comprising 35-50% of these neoplasms [95]. The presence of latent EBV in BL cells has been shown to promote genetic instability [54], suggesting a mechanism by which latent EBV could contribute to genetic alterations required for the development of BL. Other studies have shown that the priming of circulating EBV-specific CD8+ T cells is dependent on CD4+ T cells [81]. In HIV-infected individuals the CD4+ T-cell count is greatly reduced, leading to diminished CD8+ T-cell activity which permits reactivation of EBV-infected B cells [81]. High frequency of EBV association has been shown in HL (80%-100%) tissues from HIV-infected people. Additionally, EBV-encoded LMP-1 is expressed in all HIV-HL cases [86, 89], which shows that HL in HIV-infected persons appears to be an EBV-driven lymphoma [20].

Immunoblastic DLBCL has a high frequency of EBV positivity (80–90%) with frequent expression of LMP-1 and EBNA-2. LMP-1 plays a crucial role in the transformation of B-lymphocytes by EBV into immortalized human primary B cells [29] by activating NF- $\kappa\beta$ and other antiapoptotic factors [66,108].

DLBCL also gives rise to the **primary central nervous system lymphoma** (PCNSL) in AIDS patients [68]. PCNSL is extremely rare in the general population but occurs in 0.5% of patients with AIDS and shows EBV association in 100% of cases [70].

Most cases of **primary effusion lymphomas** (PELs) are dually infected with EBV and Kaposi's sarcoma associated herpesvirus (KSHV) [75]. While PEL is a rare tumor in immunocompetent individuals, it arises with increased incidence rates in HIV-infected patients. The exact role of EBV is unclear but the fact that both viruses are detected together in most cases suggests that EBV may act as a cofactor in the initiating events (because it can immortalize and transform B cells in vitro) whereas HHV-8 may be the driving force for the tumor [92].

Post-transplant lymphoproliferative disease

EBV has also been linked to most post-transplant lymphoproliferative disease (PTLD) cases, with a near 100% association. Type III latency is exhibited by the EBV-positive B cells in PTLD [15]. The wide expression of the latent EBVencoded proteins suggests an important role that EBV play in the oncogenic process. The mechanism by which EBV is thought to contribute to the pathogenesis of PTLD is similar to its presumed role in HL. As approximately 50% of PTLD cases are derived from GC B cells lacking a functional BCR because of certain crippling mutations, and because these cells manage to escape apoptosis despite lacking antigen affinity, it is believed that EBV aids in rescuing these cells from apoptosis [18, 102]. PTLD is a spectrum of lymphoid hyperproliferative states that may be observed in solid organ and bone marrow transplant recipients [69]. The majority of cases of posttransplant lymphoma (PTL) are EBV-positive [98]. Iatrogenic immunosuppression leading to primary EBV infection or reactivation of latent EBV infection is followed by polyclonal expansion of B cells. These cells are susceptible to molecular aberrations driving malignant growth [63]. Inadequate T-cell control of EBV-infected B lymphocytes is thought to be critical to the development of EBV-positive PTLDs. Decreased anti-EBV nuclear antigen antibody levels have been associated with an increased risk for PTLD [87]. Other predisposing factors include EBV mismatch between donor and recipient, high levels of immune suppression, transplanted organ type and cytomegalovirus infection [69,96].

However, there is no consensus whether any particular immunosuppressive agent is particularly responsible for PTLD. It seems that the risk of PTLD is rather low in patients treated with prednisone and azathioprine while the data on muromonab-CD3 and various forms of anti-thymocyte globulins suggest they probably increase the risk of PTLD. PTLD is commonest in children under 10 years and adults over 60 years, reaching the

REFERENCES

[1] Adhikary D., Behrends U., Moosmann A., Witter K., Bornkamm G.W., Mautner J.: Control of Epstein-Barr virus infection in vitro by T helper cells specific for virion glycoproteins. J. Exp. Med., 2006; 203: 995-1006

[2] Alexander F.E., Jarrett R.F., Lawrence D., Armstrong A.A., Freeland J., Gokhale D.A., Kane E., Taylor G.M., Wright D.H., Cartwright R.A.: Risk factors for Hodgkin's disease by Epstein-Barr virus (EBV) status: prior infection by EBV and other agents. Br. J. Cancer, 2000; 82: 1117-1121

[3] Allday M.J.: How does Epstein-Barr virus (EBV) complement the activation of myc in the pathogenesis of Burkitt's lymphoma? Semin. Cancer. Biol., 2009; 19: 366-376

[4] Allen U., Alfieri C., Preiksaitis J., Humar A., Moore D., Tapiero B., Tellier R., Green M., Davies D., Hébert D., Weitzman S., Petric M., Jacobson K.: Canadian PTLD Workshop Group - 1999. Epstein-Barr virus infection in transplant recipients: Summary of a workshop on surveillance, prevention and treatment. Can. J. Infect. Dis., 2002; 13: 89-99

[5] Arbach H., Viglasky V., Lefeu F., Guinebretière J.M., Ramirez V., Bride N., Boualaga N., Bauchet T., Peyrat J.P., Mathieu M.C., Mourah S., Podgorniak M.P., Seignerin J.M., Takada K., Joab I.: Epstein-Barr Virus (EBV) genome and expression in breast cancer tissue: effect of EBV infection of breast cancer cells on resistance to paclitaxel (Taxol). J. Virol., 2006; 80: 845-853

[6] Aydin G.B., Akyuz C., Talim B., Evans S.E., Sahin S., Sari N., Tabanlioglu D., Ozen S., Cağlar M., Büyükpamukçu M.: Extranodal type T/ NK-cell lymphoma with an atypical clinical presentation. Pediatr. Hematol. Oncol., 2007; 24: 291-299

[7] Barin F.: Viruses and unconventional transmissible agents: update on transmission via blood. Transfus. Clin. Biol., 2000; 7: 5s-10s

[8] Bevan M.J.: Helping the CD8+ T cell response. Nat. Rev. Immunol., 2004; 4: 595-602

[9] Bocian J., Januszkiewicz-Lewandowska D.: Zakażenia EBV – cykl życiowy, metody diagnostyki, chorobotwórczość. Postępy Hig. Med. Dośw., 2011; 65: 286-298 highest frequency in pediatric small bowel transplant recipients and the lowest in adult liver transplant recipients (Table 1) [79].

Table 1. Frequency of PTLD in patients after different organ transplantations by age [116].

Organ	Adults (%)	Pediatric (%)
Kidney	1.0-2.3	1.2-10.1
Liver	1.0-2.8	4.0-15.0
Heart/lung	2.4-5.8	6.4-19.5
Lung	4.2-10.0	6.4-19.5
Heart	1.0-6.3	6.4-19.5

The cumulative incidence of PTLD in allogeneic hematopoietic stem cell transplantation (HSCT) recipients is 1.0% (range 0.5-1.8%). Major risk factors for the early development of PTLD in allo-HSCT recipients include the use of unrelated or HLA-mismatched related donors, Tcell depletion of donor marrow, and use of anti-thymocyte globulin or monoclonal anti-T-cell antibodies for the prophylaxis and treatment of acute graft-versus-host disease [106].

[10] Borza C.M., Hutt-Fletcher L.M.: Alternate replication in B cells and epithelial cells switches tropism of Epstein-Barr virus. Nat. Med., 2002; 8: 594-599

[11] Boshoff C., Weiss R.: AIDS-related malignancies. Nat. Rev. Cancer, 2002; 2: 373-382

[12] Bouffet E., Frappaz D., Pinkerton R., Favrot M., Philip T.: Burkitt's lymphoma: a model for clinical oncology. Eur. J. Cancer, 1991; 27: 504-509

[13] Brady G., MacArthur G.J., Farrell P.J.: Epstein-Barr virus and Burkitt lymphoma. Postgrad. Med. J., 2008; 84: 372-377

[14] Bräuninger A., Spieker T., Willenbrock K., Gaulard P., Wacker H.H., Rajewsky K., Hansmann M.L., Küppers R.: Survival and clonal expansion of mutating "forbidden" (immunoglobulin receptor-deficient) Epstein-Barr virus-infected b cells in angioimmunoblastic t cell lymphoma. J. Exp. Med., 2001; 194: 927-940

[15] Brink A.A., Dukers D.F., van den Brule A.J., Oudejans J.J., Middeldorp J.M., Meijer C.J., Jiwa M.: Presence of Epstein- Barr virus latency type III at the single cell level in posttransplantation lymphoproliferative disorders and AIDS related lymphomas. J. Clin. Pathol., 1997; 50: 911-918

[16] Cann A.J.: Pathogenesis. In: Principles of Molecular Virology, 4th Edition, eds.: Cann A.J., Elsevier Inc, 2005, 238

[17] Capello D., Rasi S., Oreste P., Veronese S., Cerri M., Ravelli E., Rossi D., Minola E., Colosimo A., Gambacorta M., Muti G., Morra E., Gaidano G.: Molecular characterization of post-transplant lymphoproliferative disorders of donor origin occurring in liver transplant recipients. J. Pathol., 2009; 218: 478-486

[18] Capello D., Rossi D., Gaidano G.: Post-transplant lymphoproliferativedisorders: molecular basis of disease histogenesis and pathogenesis. Hematol. Oncol., 2005; 23: 61-67

[19] Carbone A., Gloghini A., Dotti G.: EBV-associated lymphoproliferative disorders: classification and treatment. Oncologist, 2008; 13: 577-585 [20] Carbone A., Gloghini A., Larocca L.M., Antinori A., Falini B., Tirelli U., Dalla-Favera R., Gaidano G.: Human immunodeficiency virus-associated Hodgkin's disease derives from post-germinal center B cells. Blood, 1999; 93: 2319-2326

[21] Cartwright R.A., Watkins G.: Epidemiology of Hodgkin's disease: a review. Hematol. Oncol., 2004; 22: 11-26

[22] Cesarman E., Mesri E.A.: Virus associated lymphomas. Curr. Opin. Oncol., 1999; 11: 322-332

[23] Chan K.H., Ng M.H., Seto W.H., Peiris J.S.: Epstein-Barr virus (EBV) DNA in sera of patients with primary EBV infection. J. Clin. Microbiol., 2001; 39: 4152-4154

[24] Chapman A.L., Rickinson A.: Epstein-Barr virus in Hodgkin's disease. Ann. Oncol., 1998; 9: S5-S16

[25] Chen M.R.: Epstein-Barr virus, the immune system, and associated diseases. Front Microbiol. 2011; 2: 5

[26] Cohen J.I., Bollard C.M., Khanna R., Pittaluga S.: Current understanding of the role of Epstein-Barr virus (EBV) in lymphomagenesis and therapeutic approaches to EBV-associated lymphomas. Leuk. Lymphoma, 2008; 49: 27-34

[27] de Leval L., Hasserjian R.P.: Diffuse large B cell lymphomas and burkitt lymphoma. Hematol. Oncol. Clin. North Am., 2009; 23: 791-827

[28] Deacon E.M., Pallesen G., Niedobitek G., Crocker J., Brooks L., Rickinson A.B., Young L.S.: Epstein-Barr virus and Hodgkin's disease: transcriptional analysis of virus latency in the malignant cells. J. Exp. Med., 1993; 177: 339-349

[29] Dirmeier U., Neuhierl B., Kilger E., Reisbach G., Sandberg M.L., Hammerschmidt W.: Latent membrane protein 1 is critical for efficient growth transformation of human B cells by Epstein-Barr virus. Cancer Res., 2003; 63: 2982-2989

[30] Dorsett Y., Robbiani D.F., Jankovic M., Reina-San-Martin B., Eisenreich T.R., Nussenzweig M.C.: A role for AID in chromosome translocations between c-myc and the IgH variable region. J. Exp. Med., 2007; 204: 2225-2232

[31] Eligio P., Delia R., Valeria G.: EBV chronic infections. Mediterr. J. Hematol. Infect. Dis., 2010; 2: e2010022

[32] Epstein M.A., Achong B.G., Barr Y.M.: Virus particles in cultured lymphoblasts from Burkitt's lymphoma. Lancet, 1964; 1: 702-703

[33] Farrell K., Jarrett R.F.: The molecular pathogenesis of Hodgkin lymphoma. Histopathology, 2011; 58: 15-25

[34] Filipovich A.H., Mathur A., Kamat D., Shapiro R.S.: Primary immunodeficiencies: genetic risk factors for lymphoma. Cancer Res., 1992; 52: 5465s-5467s

[35] Fingeroth J.D., Weis J.J., Tedder T.F., Strominger J.L., Biro P.A., Fearon D.T.: Epstein-Barr virus receptor of human B-lymphocytes in the C3d receptor CR2. Proc. Natl. Acad. Sci. USA, 1984; 81: 4510-4514

[36] God J.M., Haque A.: Burkitt lymphoma: pathogenesis and immune evasion. J. Oncol., 2010; 2010: 516047

[37] Gratama J.W., Ernberg I.: Molecular epidemiology of Epstein-Barr virus infection. Adv. Cancer Res., 1995; 67: 197-255

[38] Harris N.L., Jaffe E.S., Diebold J., Flandrin G., Muller-Hermelink H.K., Vardiman J., Lister T.A., Bloomfield C.D.: The World Health Organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues. Report of the Clinical Advisory Committee meeting, Airlie House, Virginia, November, 1997. Ann. Oncol., 1999; 10: 1419-1432

[39] Herbst H., Foss H.D., Samol J., Araujo I., Klotzbach H., Krause H., Agathanggelou A., Niedobitek G., Stein H.: Frequent expression of interleukin-10 by Epstein-Barr virus-harboring tumor cells of Hodgkin's disease. Blood, 1996; 87: 2918-2929

[40] Higgins C.D., Swerdlow A.J., Macsween K.F., Harrison N., Williams H., McAulay K., Thomas R., Reid S., Conacher M., Britton K., Crawford D.H.: A study of risk factors for acquisition of Epstein-Barr virus and its subtypes. J. Infect. Dis., 2007; 195: 474-482

[41] Hislop A.D., Taylor G.S., Sauce D., Rickinson A.B.: Cellular responses to viral infection in humans: lessons from Epstein-Barr virus. Annu. Rev. Immunol., 2007; 25: 587-617

[42] Hjalgrim H., Friborg J., Melbye M.: The epidemiology of EBV and its association with malignant disease. In: Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis, eds., A. Arvin, G. Campadelli-Fiume, E. Mocarski, P.S. Moore, B. Roizman, R. Whitley, K. Yamanishi. Cambridge University Press 2007, 929-959

[43] Hodgkin lymphoma. In: WHO Classification of Tumors: Pathology and Genetics of Tumors of Haematopoietic and Lymphoid Tissues, vol.3, eds.: S.H. Swerdlow, E. Campo, N.L. Harris, E.S. Jaffe, S.A. Pileri, S. Harald, J. Thiele, J.W. Vardiman. IARC Press, Lyon 2008, 321-334

[44] Honjo T., Kinoshita K., Muramatsu M.: Molecular mechanism of class switch recombination: linkage with somatic hypermutation. Annu. Rev. Immunol., 2002; 20: 165-196

[45] Hsu S.M., Lin J., Xie S.S., Hsu P.L., Rich S.: Abundant expression of transforming growth factor-beta 1 and -beta 2 by Hodgkin's Reed--Sternberg cells and by reactive T lymphocytes in Hodgkin's disease. Hum. Pathol., 1993; 24: 249-255

[46] Iannitto E., Ferreri A.J., Minardi V., Tripodo C., Kreipe H.H.: Angioimmunoblastic T cell lymphoma. Crit. Rev. Oncol. Hematol., 2008; 68: 264-271

[47] IARC Monographs on the evaluation of carcinogenic risks to humans, vol. 70: Epstein-Barr virus and Kaposi's sarcoma herpesvirus/herpesvirus. World Health Organization, International Agency for Research on Cancer. IARC Press, Lyon 1997

[48] Jaccard A., Hermine O.: Extranodal natural killer/T cell lymphoma: advances in the management. Curr. Opin. Oncol., 2011; 23: 429-435

[49] Jaffe E.S., Chan J.K., Su IJ, Frizzera G., Mori S., Feller A.C., Ho F.C.: Report of the workshop on nasal and related extranodal angiocentric T/natural killer cell lymphomas. Am. J. Surg. Pathol., 1996; 20: 103-111

[50] Jaffe E.S., Ralfkiaer E.: Angioimmunoblastic T-cell lymphoma. In:. Pathology and Genetics: Tumours of Haematopoietic and Lymphoid Tissues, eds.: E.S. Jaffe, N.L. Harris, H. Stein, J.W. Vardiman. IRAC Press, Lyon 2001, 225-226

[51] Jarrett A.F., Armstrong A.A., Alexander E.: Epidemiology of EBV and Hodgkin's lymphoma. Ann. Oncol., 1996; 7: 5-10

[52] Jones J.F., Shurin S., Abramowsky C., Tubbs R.R., Sciotto C.G., Wahl R., Sands J., Gottman D., Katz B.Z., Sklar J.: T cell lymphomas containing Epstein-Barr virus DNA in patients with chronic Epstein-Barr virus infection. N. Engl. J. Med., 1988; 318: 733-741

[53] Joo E.J., Ha Y.E., Jung D.S., Cheong H.S., Wi Y.M., Song J.H., Peck K.R.: An adult case of chronic active Epstein-Barr virus infection with interstitial pneumonitis. Korean J. Intern. Med., 2011; 26: 466-469

[54] Kamranvar S.A., Gruhne B., Szeles A., Masucci M.G.: Epstein-Barr virus promotes genomic instability in Burkitt's lymphoma. Oncogene, 2007; 26: 5115-5123

[55] Kanegane H., Bhatia K., Gutierrez M., Kaneda H., Wada T., Yachie A., Seki H., Arai T., Kagimoto S., Okazaki M., Oh-ishi T., Moghaddam A., Wang F., Tosato G.: A syndrome of peripheral blood T cell infection with Epstein Barr virus (EBV) followed by EBV-positive T cell lymphoma. Blood, 1998; 91: 2085-2091

[56] Kanegane H., Nomura K., Miyawaki T., Tosato G.: Biological aspects of Epstein-Barr virus (EBV)-infected lymphocytes in chronic active EBV infection and associated malignancies. Crit. Rev. Oncol. Hematol., 2002; 44: 239-249 [57] Kapatai G., Murray P.: Contribution of the Epstein Barr virus to the molecular pathogenesis of Hodgkin lymphoma. J. Clin. Pathol., 2007; 60: 1342-1349

[58] Kapp U., Yeh W.C., Patterson B., Elia A.J., Kägi D., Ho A., Hessel A., Tipsword M., Williams A., Mirtsos C., Itie A., Moyle M., Mak TW.: Interleukin 13 is secreted by and stimulates the growth of Hodgkin and Reed-Sternberg cells. J. Exp. Med., 1999; 189: 1939-1946

[59] Kawa-Ha K., Ishihara S., Ninomiya T., Yumura-Yagi K., Hara J., Murayama F., Tawa A., Hirai K.: CD3-negative lymphoproliferative disease of granular lymphocytes containing Epstei-Barr viral DNA. J. Clin. Invest., 1989; 84: 51-55

[60] Kelly G.L., Milner A.E., Baldwin G.S., Bell A.I., Rickinson A.B.: Three restricted forms of Epstein-Barr virus latency counteracting apoptosis in c-myc–expressing Burkitt lymphoma cells. Proc. Natl. Acad. Sci. USA, 2006; 103: 14935-14940

[61] Khan G., Miyashita E.M., Yang B., Babcock G.J., Thorley-Lawson D. A.: Is EBV persistence in vivo a model for B cell homeostasis? Immunity, 1996; 5: 173-179

[62] Kitagawa N., Goto M., Kurozumi K., Maruo S., Fukayama M., Naoe T., Yasukawa M., Hino K., Suzuki T., Todo S., Takada K.: Epstein-Barr virus-encoded poly(A)(-) RNA supports Burkitt's lymphoma growth through interleukin-10 induction. EMBO J., 2000; 19: 6742-6750

[63] Knowles D.M.: The molecular genetics of post-transplantation lymphoproliferative disorders. Springer Semin. Immunopathol., 1998; 20: 357-373

[64] Knowles D.: Etiology and pathogenesis of AIDS-related non-Hodgkin's lymphoma. Hematol. Oncol. Clin. N. Am., 2003; 17: 785-820

[65] Küppers R.: B cells under influence: transformation of B cells by Epstein–Barr virus. Nat. Rev. Immunol., 2003; 3: 801-812

[66] Kulwichit W., Edwards R.H., Davenport E.M., Baskar J.F., Godfrey V., Raab-Traub N.: Expression of the Epstein-Barr virus latent membrane protein 1 induces B cell lymphoma in transgenic mice. Proc. Natl. Acad. Sci. USA, 1998; 95: 11963-11968

[67] Kutok J.L., Wang F.: Spectrum of Epstein-Barr virus-associated diseases. Annu. Rev. Pathol., 2006; 1: 375-404

[68] Levine A.M.: Acquired immunodeficiency syndrome-related lymphoma. Blood, 1992; 80: 8-20

[69] Loren A.W., Porter D.L., Stadtmauer E.A., Tsai D.E.: Post-transplant lymphoproliferative disorder: a review. Bone Marrow Transplant., 2003; 31: 145-155

[70] MacMahon E.M., Glass J.D., Hayward S.D., Mann R.B., Charache P., McArthur J.C., Ambinder R.F.: Association of Epstein-Barr virus with primary central nervous system lymphoma in AIDS. AIDS Res. Hum. Retroviruses, 1992; 8: 740-742

[71] Mueller N., Evans A., Harris N.L., Comstock G.W., Jellum E., Magnus K., Orentreich N., Polk B.F., Vogelman J.: Hodgkin's disease and Epstein-Barr virus. Altered antibody pattern before diagnosis. N. Engl. J. Med., 1989; 320: 689-695

[72] Münz C., Moormann A.: Immune escape by Epstein Barr virus associated malignancies, Semin. Cancer Biol., 2008; 18: 381-387

[73] Murray P.G., Young L.S.: The role of the Epstein-Barr virus in human disease. Front Biosci., 2002; 7: d519-d540

[74] Mutalima N., Molyneux E., Jaffe H., Kamiza S., Borgstein E., Mkandawire N., Liomba G., Batumba M., Lagos D., Gratrix F., Boshoff C., Casabonne D., Carpenter L.M., Newton R.: Associations between Burkitt lymphoma among children in Malawi and infection with HIV, EBV and malaria: results from a case-control study. PLoS One, 2008; 3: e2505

[75] Nador R.G., Cesarman E., Chadburn A., Dawson D.B., Ansari M.Q., Sald J., Knowles D.M.: Primary effusion lymphoma: a distinct clinicopathologic entity associated with the Kaposi's sarcoma-associated herpesvirus. Blood, 1996; 88: 645-656 [76] Nikiforow S., Bottomly K., Miller G., Münz C.: Cytolytic CD4+-T cell clones reactive to EBNA-1 inhibit Epstein-Barr virus-induced B cell proliferation. J. Virol., 2003; 77:12088-12104

[77] Nilsson K.: Human B-lymphoid cell lines. Hum. Cell, 1992; 5: 25-41

[78] Ong K.W., Teo M., Lee V., Ong D., Lee A., Tan C.S., Vathsala A., Toh H.C.: Expression of EBV latent antigens, mammalian target of rapamycin, and tumor suppression genes in EBV-positive smooth muscle tumors: clinical and therapeutic implications. Clin. Cancer Res., 2009; 15: 5350-5358

[79] Parker A., Bowles K., Bradley J.A., Emery V., Featherstone C., Gupte G., Marcus R., Parameshwar J., Ramsay A., Newstead C.: Diagnosis of post-transplant lymphoproliferative disorder in solid organ transplant recipients – BCSH and BTS guidelines. Br. J. Haematol., 2010; 149; 675-692

[80] Pasqualucci L., Bhagat G., Jankovic M., Compagno M., Smith P., Muramatsu M., Honjo T., Morse H.C., Nussenzweig M.C., Dalla-Favera R.: AID is required for germinal center-derived lymphomagenesis. Nat. Genet., 2008; 40: 108-112

[81] Pietersma F., Piriou E., van Baarle D.: Immune surveillance of EBV--infected B cells and the development of non-Hodgkin lymphomas in immunocompromised patients. Leuk. Lymphoma, 2008; 49: 1028-1041

[82] Quintanilla-Martinez L., Kumar S., Fend F., Reyes E., Teruya-Feldstein J., Kingma D.W., Sorbara L., Raffeld M., Straus S.E., Jaffe E.S.: Fulminat EBV (+) T cell lymphoproliferative disorder following acute/chronic EBV infection: a distinct clinicopathologic syndrome. Blood, 2000; 96: 443–451

[83] Raab-Traub N.: EBV-induced oncogenesis. In: Human Herpesviruses: Biology, Therapy, and Immunoprophylaxis, eds.: A. Arvin, G. Campadelli-Fiume, E. Mocarski, P.S. Moore, B. Roizman, R. Whitley, K. Yamanishi. Cambridge University Press, Cambridge 2007, 986-1006

[84] Raphael M., Borisch B., Jaffe E.: Lymphomas associated with infection by the human immune deficiency virus (HIV). In: World Health Organization classification of tumors, pathology and genetics of tumors of haematopoietic and lymphoid tissues, eds.L Jaffe E., Harris N., Stein H., Vardiman J. IARC Press, Lyon, 2001, 260-263

[85] Re D., Küppers R., Diehl V.: Molecular pathogenesis of Hodgkin's lymphoma. J. Clin. Oncol., 2005; 23: 6379-6386

[86] Rezk S.A., Weiss L.M.: Epstein-Barr virus-associated lymphoproliferative disorders. Hum. Pathol., 2007; 38: 1293-1304

[87] Riddler S.A., Breinig M.C., McKnight J.L.: Increased levels of circulating Epstein-Barr virus (EBV)-infected lymphocytes and decreased EBV nuclear antigen antibody responses are associated with the development of posttransplant lymphoproliferative disease in solid-organ transplant recipients. Blood, 1994; 84: 972-984

[88] Rizvi M.A., Evens A.M., Tallman M.S., Nelson B.P., Rosen S.T.: T cell non-Hodgkin's lymphoma. Blood, 2006; 107: 1255-1264

[89] Said J.W.: Immunodeficiency-related Hodgkin lymphoma and its mimics. Adv. Anat. Pathol.; 2007; 14: 189-194

[90] Sample J., Young L., Martin B., Chatman T., Kieff E., Rickinson A., Kieff E.: Epstein-Barr virus types 1 and 2 differ in their EBNA-3A, EBNA-3B, and EBNA-3C genes. J. Virol., 1990; 64: 4084-4092

[91] Sbih-Lammali F., Djennaoui D., Belaoui H., Bouguermouh A., Decaussin G., Ooka T.: Transcriptional expression of Epstein-Barr virus genes and proto-oncogenes in north African nasopharyngeal carcinoma. J. Med. Virol., 1996; 49: 7-14

[92] Simonelli C., Spina M., Cinelli R., Talamini R., Tedeschi R., Gloghini A., Vaccher E., Carbone A., Tirelli U.: Clinical features and outcome of primary effusion lymphoma in HIV-infected patients: a single--institution study. J. Clin. Oncol., 2003; 21: 3948-3954

[93] Sixbey J.W., Nedrud J.G., Raab-Traub N., Hanes R.A., Pagano J.S.: Epstein-Barr virus replication in oropharyngeal epithelial cells. N. Engl. J. Med., 1984; 310: 1225-1230 [94] Sixbey J.W., Shirley P., Chesney P.J., Buntin D.M., Resnick L.: Detection of a second widespread strain of Epstein-Barr virus. Lancet, 1989; 2: 761-765

[95] Spina M., Tirelli U., Zagonel V., Gloghini A., Volpe R., Babare R., Abbruzzese L., Talamini R., Vaccher E., Carbone A.: Burkitt's lymphoma in adults with and without human immunodeficiency virus infection: a single-institution clinicopathologic study of 75 patients. Cancer, 1998; 82: 766-774

[96] Swinnen L.J.: Posttransplant lymphoproliferative disorders. In: Infectious causes of cancer: Targets for intervention, eds.: J.J. Goedert. Humana Press, Totowa 2000, 63-76

[97] Szmidt A., Stańczyk-Przyłuska A.: Rola wirusa EBV w patogenezie chorób alergicznych. Alerg. Astma Immunol., 2005; 10: 169-174

[98] Taylor A..L, Marcus R., Bradley J.A.: Post-transplant lymphoproliferative disorders (PTLD) after solid organ transplantation. Crit. Rev. Oncol. Hematol., 2005; 56: 155-167

[99] Thompson M.P., Kurzrock R.: Epstein-Barr virus and cancer. Clin. Cancer Res., 2004; 10: 803-821

[100] Thompson S., Messick T., Schultz D.C., Reichman M., Lieberman P.M.: Development of a high-throughput screen for inhibitors of Epstein-Barr virus EBNA1. J. Biomol. Screen., 2010; 15: 1107-1115

[101] Thorley-Lawson D.A.: Epstein-Barr virus: exploiting the immune system. Nat. Rev. Immunol., 2001; 1: 75-82

[102] Timms J.M., Bell A., Flavell J.R., Murray P.G., Rickinson A.B., Traverse-Glehen A., Berger F., Delecluse H.J.: Target cells of Epstein-Barr-virus (EBV)-positive post-transplant lymphoproliferative disease: similarities to EBV-positive Hodgkin's lymphoma. Lancet, 2003; 361: 217-223

[103] Toczyski D.P., Matera A.G., Ward D.C., Steitz J.A.: The Epstein-Barr virus (EBV) small RNA EBER1 binds and relocalizes ribosomal protein L22 in EBV-infected human B lymphocytes. Proc. Natl. Acad. Sci. USA, 1994; 91: 3463-3467

[104] Trottier H., Buteau C., Robitaille N., Duval M., Tucci M., Lacroix J., Alfieri C.: Transfusion-related Epstein-Barr virus infection among stem cell transplant recipients: a retrospective cohort study in children. Transfusion, 2012; 52: 2653-2663

[105] van den Bosch C.A.: Is endemic Burkitt's lymphoma an alliance between three infections and a tumor promoter? Lancet Oncol., 2004; 5: 738-746

[106] Wagner H.J., Rooney C.M., Heslop H.E.: Diagnosis and treatment of posttransplantation lymphoproliferative disease after hematopoietic stem cell transplantation. Biol. Blood Marrow Transplant., 2002; 8: 1-8 [107] Wakiguchi H.: Overview of Epstein-Barr virus-associated diseases in Japan. Crit Rev. Oncol. Hematol., 2002; 44: 193-202

[108] Wang D., Liebowitz D., Kieff E.: An EBV membrane protein expressed in immortalized lymphocytestransforms established rodent cells. Cell, 1985; 43: 831-840

[109] Weiss L.M., Jaffe E.S., Liu X.F., Chen Y.Y., Shibata D., Medeiros L.J.: Detection and localization of Epstein-Barr viral genomes in angioimmunoblastic lymphadenopathy and angioimmunoblastic lymphadenopathy-like lymphoma. Blood, 1992; 79: 1789-1795

[110] Weiss L.M., Strickler J.G., Warnke R.A., Purtilo D.T., Sklar J.: Epstein-Barr viral DNA in tissues of Hodgkin's disease. Am. J. Pathol., 1987; 129: 86-91

[111] Wekerle H., Hohlfeld R.: Molecular mimicry in multiple sclerosis. N. Engl. J. Med., 2003; 349: 185-186

[112] Wensing B., Farrell P.J.: Regulation of cell growth and death by Epstein-Barr virus. Microbes Infect., 2000; 2: 77-84

[113] Went P., Agostinelli C., Gallamini A., Piccaluga P.P., Ascani S., Sabattini E., Bacci F., Falini B., Motta T., Paulli M., Artusi T., Piccioli M., Zinzani P.L., Pileri S.A.: Marker expression in peripheral T cell lymphoma: A proposed clinical-pathologic prognostic score. J. Clin. Oncol., 2006; 24: 2472-2479

[114] Wilson J.B., Bell J.L., Levine A.J.: Expression of Epstein-Barr virus nuclear antigen-1 induces B cell neoplasia in transgenic mice. EMBO J., 1996; 15: 3117-3126

[115] Wright D.H.: What is Burkitt's lymphoma and when is it endemic? Blood, 1999; 93: 758

[116] Young L.S.: Epstein-Barr Virus: General Features. In: Encyclopedia of Virology, eds.: Mahy B.W.J., van Regenmortel M.H.V., Academic Press, Oxford 2008,148-157

[117] Young L.S., Murray P.G.: Epstein-Barr virus and oncogenesis: from latent genes to tumors. Oncogene, 2003; 22, 5108-5121

[118] Young L.S., Rickinson A.B.: Epstein-Barr virus: 40 years on. Nat. Rev. Cancer, 2004; 4: 757-768

[119] Yustein J.T., Dang C.V.: Biology and treatment of Burkitt's lymphoma. Curr. Opin. Hematol., 2007; 14: 375-381

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